

# Galaxy 101

## Overview

### Questions

- Which coding exon has the highest number of single nucleotide polymorphisms (SNPs) on human chromosome 22?

### Objectives

- Familiarize yourself with the basics of Galaxy
- Learn how to obtain data from external sources
- Learn how to run tools
- Learn how histories work
- Learn how to create a workflow
- Learn how to share your work

 **Time estimation:** 1-1.5h

## 101 Introduction

This practical aims to familiarize you with the Galaxy user interface. It will teach you how to perform basic tasks such as importing data, running tools, working with histories, creating workflows, and sharing your work.

## Agenda

In this tutorial, we will:

1. Pretreatments
  1. Upload exon locations
  2. Upload SNP information
2. Analysis
  1. Find exons with the highest number of SNPs
  2. Count the number of SNPs per exon
  3. Sort the exons by SNPs count
  4. Select the top five exons
  5. Recovering exon info and displaying data in genome browsers
  6. UCSC genome browser
3. Galaxy management
  1. Convert your analysis history into a workflow
  2. The workflow editor
  3. Run workflow on different data
  4. Share your work

## Pretreatments

Suppose you get the following question:

### ?

#### Question

*Mom (or Dad) ... Which coding exon has the highest number of single nucleotide polymorphisms (SNPs) on human chromosome 22?*

You are thinking "Wow! This is a simple question... I know where to find the data, at the UCSC Genome Browser (<https://genome.ucsc.edu/>), but how do I actually compute this?" There is really a straightforward way of answering this question and it is called **Galaxy**. So let's try it...

Browse to your Galaxy instance and log in or register. The Galaxy interface consists of three main parts. The available tools are listed on the left, your analysis history is recorded on the right, and the middle pane will show the home page, the selected tool or the dataset.

Galaxy is an open source, web-based platform for data intensive biomedical research. If you are new to Galaxy [start here](#) or consult our [help resources](#). You can install your own Galaxy by following the [tutorial](#) and choose from thousands of tools from the [Tool Shed](#).

**Running Your Own Understanding how Galaxy works**  
An in-depth tutorial

**Main**

**History**

The Galaxy Team is a part of the Center for Comparative Genomics and Bioinformatics at Penn State, the Department of Biology and at Johns Hopkins University and the Computational Biology Program at Oregon Health & Science University.

This instance of Galaxy is utilizing infrastructure generously provided by the CyVerse at the Texas Advanced Computing Center, with support from the National Science Foundation.

**Tools**

- Get Data
- Send Data
- Lift-Over
- Collection Operations
- Text Manipulation
- Datamash
- Convert Formats
- Filter and Sort
- Join, Subtract and Group
- Fetch Alignments/Sequences
- NGS: QC and manipulation
- NGS: DeepTools
- NGS: Mapping
- NGS: RNA Analysis
- NGS: SAMTools
- NGS: BamTools
- NGS: Picard
- NGS: VCF Manipulation
- NGS: Peak Calling
- NGS: Variant Analysis
- NGS: RNA Structure
- NGS: Du Novo
- NGS: Gemini
- NGS: Assembly
- NGS: Chromosome Conformation
- NGS: Mothur
- Operate on Genomic Intervals
- Statistics
- Graph/Display Data
- Phenotype Association
- BEDTools
- Genome Diversity
- EMBOSS
- Regional Variation
- FASTA manipulation
- Multiple Alignments
- Metagenomic Analysis



## Hands-on: Create history

1. Make sure you have an empty analysis history.



### Starting a new history

- o Click the **gear icon** at the top of the history panel
- o Select the option **Create New** from the menu

2. **Rename your history** to be meaningful and easy to find. You can do this by clicking on the title of the history (by default the title is *Unnamed history*) and typing **Galaxy 101** as the name. Do



not forget to hit **enter** on your keyboard to save it.

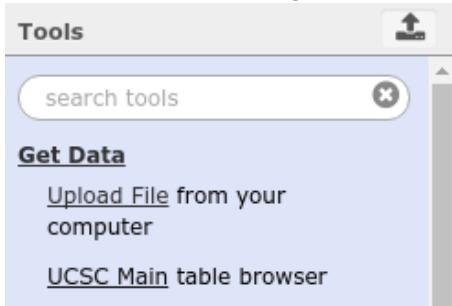
## Upload exon locations

Now we are ready to do some analysis, but first we will need to get some data into our history. You can upload files from your computer, but Galaxy can also fetch data directly from external sources. We will now import a list of all the exon locations on chromosome 22 directly from the UCSC table browser.



## Hands-on: Data upload from UCSC

1. In the tool menu, navigate to Get Data -> UCSC Main - table browser



You will be taken to the **UCSC table browser**, which looks something like this:

**Table Browser**

Use this program to retrieve the data associated with a track in text format, to calculate intersections between tracks, and to retrieve DNA sequence covered by a track. For help in using this application see [Using the Table Browser](#) for a description of the controls in this form, the [User's Guide](#) for general information and sample queries, and the OpenHelix Table Browser [tutorial](#) for a narrated presentation of the software features and usage. For more complex queries, you may want to use [Galaxy](#) or our [public MySQL server](#). To examine the biological function of your set through annotation enrichments, send the data to [GREAT](#). Send data to [GenomeSpace](#) for use with diverse computational tools. Refer to the [Credits](#) page for the list of contributors and usage restrictions associated with these data. All tables can be downloaded in their entirety from the [Sequence and Annotation Downloads](#) page.

**clade:** Mammal    **genome:** Human    **assembly:** Feb. 2009 (GRCh37/hg19) ▾  
**group:** Genes and Gene Predictions    **track:** UCSC Genes    [add custom tracks](#)    [track hubs](#)  
**table:** knownGene    [describe table schema](#)  
**region:**  genome  ENCODE Pilot regions  position chr22    [lookup](#)    [define regions](#)  
**identifiers (names/acceessions):** [paste list](#)    [upload list](#)  
**filter:** [create](#)  
**intersection:** [create](#)  
**correlation:** [create](#)  
**output format:** BED - browser extensible data     Send output to  [Galaxy](#)     [GREAT](#)     [GenomeSpace](#)  
**output file:**  (leave blank to keep output in browser)  
**file type returned:**  plain text  gzip compressed  
  
[get output](#)    [summary/statistics](#)

To reset all user cart settings (including custom tracks), [click here](#).



## Settings

- **clade** should be set to Mammal
- **genome** should be set to Human
- **assembly** should be set to Feb. 2009 (GRCh37/hg19)
- **group** should be set to Genes and Gene Predictions
- **region** should be changed to position with value chr22
- **output format** should be set to BED - browser extensible data
- **Send output to** should have the option Galaxy checked

2. Click on the **get output** button and you will see the next screen:

Home Genomes Genome Browser Tools Mirrors Downloads My Data Help About Us

**Output knownGene as BED**

**Include custom track header:**

name=   
 description=   
 visibility=   
 url=

**Create one BED record per:**

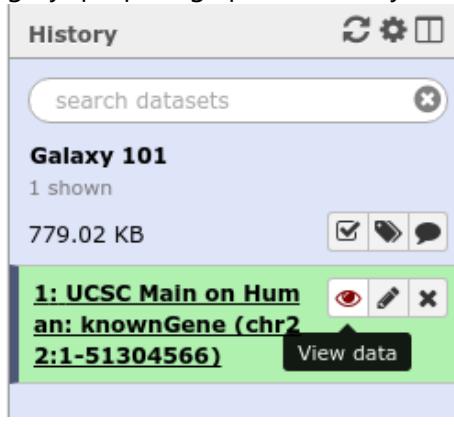
Whole Gene  
 Upstream by  bases  
 Exons plus  bases at each end  
 Introns plus  bases at each end  
 5' UTR Exons  
 Coding Exons  
 3' UTR Exons  
 Downstream by  bases

Note: if a feature is close to the beginning or end of a chromosome and upstream/downstream bases are added, they may be truncated in order to avoid extending past the edge of the chromosome.

Change **Create one BED record per** to **Coding Exons** and then click on the **Send Query to Galaxy** button.



After this you will see your first history item in Galaxy's right pane. It will go through the gray (preparing/queued) and yellow (running) states to become green (success):



3. When the dataset is green, click on the **eye icon** to **view the contents** of the file. It should look something like this:

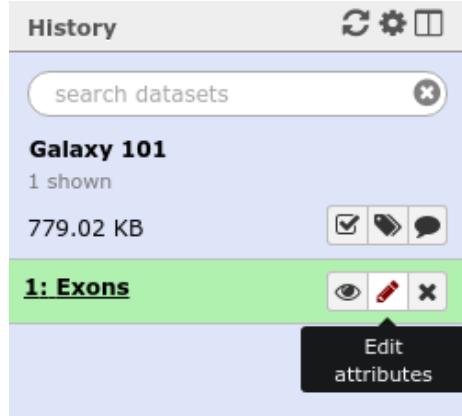
1	2	3	4	5	6
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-
chr22	16268136	16268181	uc002zlh.1_cds_3_0_chr22_16268137_r	0	-
chr22	16269872	16269943	uc002zlh.1_cds_4_0_chr22_16269873_r	0	-
chr22	16275206	16275277	uc002zlh.1_cds_5_0_chr22_16275207_r	0	-
chr22	16277747	16277885	uc002zlh.1_cds_6_0_chr22_16277748_r	0	-

Each line represents an exon, the first three columns are the genomic location, and the fourth column contains the ID (name) of the exon.

4. Let's rename our dataset to something more recognizable.

- o Click on the **pencil icon** to edit a file's attributes.
- o In the next screen change the name of the dataset to **Exons**.
- o Click the **Save** button at the bottom of the screen.

Your history should now look something like this:



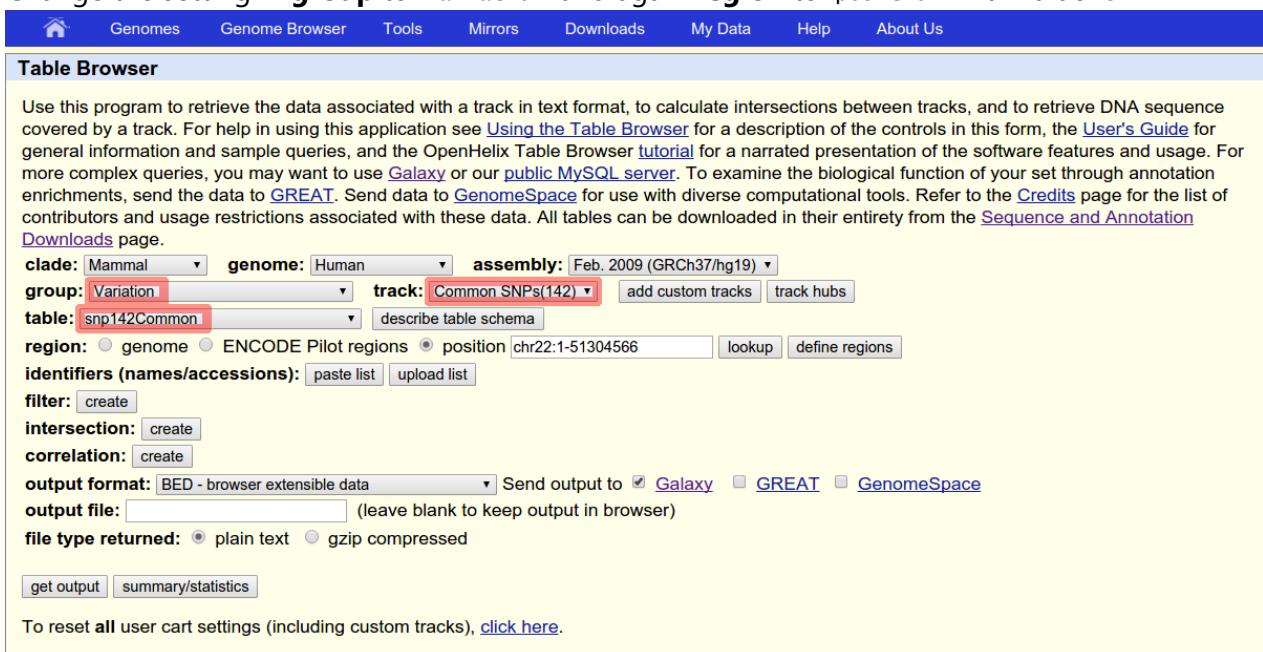
## Upload SNP information

Now we have information about the exon locations, but our question was which exon contains the largest number of SNPs, so let's get some information about SNP locations from UCSC as well:



## Hands-on: SNP information

1. **UCSC Main** : Return to the UCSC tool ucsc Main - table browser
2. Change the setting in **group** to variation and again **region** to position with value chr22



Use this program to retrieve the data associated with a track in text format, to calculate intersections between tracks, and to retrieve DNA sequence covered by a track. For help in using this application see [Using the Table Browser](#) for a description of the controls in this form, the [User's Guide](#) for general information and sample queries, and the OpenHelix Table Browser [tutorial](#) for a narrated presentation of the software features and usage. For more complex queries, you may want to use [Galaxy](#) or our [public MySQL server](#). To examine the biological function of your set through annotation enrichments, send the data to [GREAT](#). Send data to [GenomeSpace](#) for use with diverse computational tools. Refer to the [Credits](#) page for the list of contributors and usage restrictions associated with these data. All tables can be downloaded in their entirety from the [Sequence and Annotation Downloads](#) page.

clade: Mammal   genome: Human   assembly: Feb, 2009 (GRCh37/hg19)  

group: Variation   track: Common SNPs(142)   add custom tracks   track hubs

table:.snp142Common   describe table schema

region: genome   ENCODE Pilot regions   position: chr22:1-51304566   lookup   define regions

identifiers (names/accessions): paste list   upload list

filter: create

intersection: create

correlation: create

output format: BED - browser extensible data   Send output to: Galaxy   GREAT   GenomeSpace

output file: (leave blank to keep output in browser)

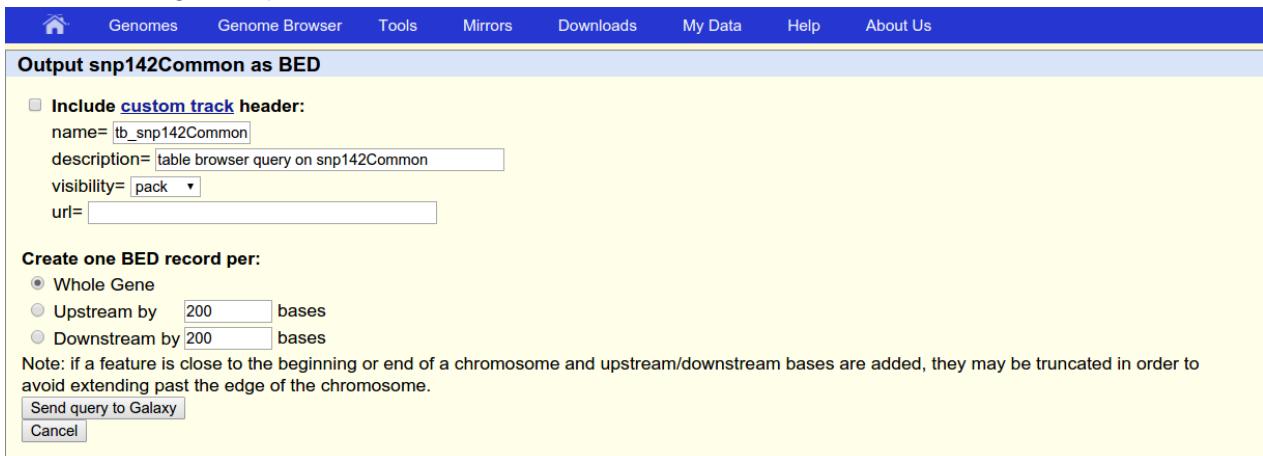
file type returned: plain text   gzip compressed

get output   summary/statistics

To reset all user cart settings (including custom tracks), [click here](#).

The **track** setting shows the version of the SNP database to get. In this example it is version 142, but you may select the latest one. Your results may vary slightly from the ones in this tutorial when you select a different version, but in general it is a good idea to select the latest version, as this will contain the most up-to-date SNP information.

3. Click on the `get output` button to find a menu similar to this:



Output.snp142Common as BED

Include [custom track header](#):

name= tb\_snp142Common  
description= table browser query on.snp142Common  
visibility= pack  
url=

Create one BED record per:

Whole Gene  
 Upstream by 200 bases  
 Downstream by 200 bases

Note: If a feature is close to the beginning or end of a chromosome and upstream/downstream bases are added, they may be truncated in order to avoid extending past the edge of the chromosome.

[Send query to Galaxy](#)   [Cancel](#)

Make sure that **Create one BED per** is set to `whole Gene` (Whole Gene here really means Whole Feature), and click on **Send Query to Galaxy**. You will get your second item in your analysis history.

4. Now **rename** your new dataset to `SNPs` so we can easily remember what the file contains.

## Analysis

### Find exons with the highest number of SNPs

Let's remind ourselves that our objective was to find which exon contains the most SNPs. Therefore we have to join the file with the exon locations with the file containing the SNP locations (here "join" is just a fancy word for printing the SNPs and exons that overlap side-by-side).

### 💡 Search bar

Different Galaxy servers may have tools available under different sections, therefore it is often useful to use the **search bar** at the top of the tool panel to find your tool.

### 📝 Hands-on: Finding Exons

1. **Join** : Enter the word `join` in the search bar of the tool panel, and select the tool named `Join - the intervals of two datasets side-by-side`
2. Select your file with exons as the first file, and the file with SNPs as the second file, and make sure **return** is set to `INNER JOIN` so that only matches are included in the output (i.e. only exons with SNPs in it and only SNPs that fall in exons)

**Note:** if you scroll down on this page, you will find the help of the tool.

3. Click the **Execute** button and view the resulting file (with the eye icon). If everything went okay, you should see a file that looks similar to this:

1	2	3	4	5	6	7	8	9	10	11	12
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-	chr22	16287850	16287851	rs72485235	0	+
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-	chr22	16287537	16287538	rs200179046	0	+
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-	chr22	16287338	16287339	rs199952431	0	+
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-	chr22	16287393	16287394	rs201714672	0	+
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-	chr22	16287345	16287346	rs200013113	0	+
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-	chr22	16287371	16287372	rs201840700	0	+

Remember that variations are possible due to using different versions of UCSC databases, as long as you have similar looking columns you did everything right :)

Let's take a look at this dataset. The first six columns correspond to the exons, and the last six columns correspond to the SNPs. Column 4 contains the exon IDs, and column 10 contains the SNP IDs. In our screenshot you see that the first 5 lines in the file all have the same exon ID ( uc010gqp.2\_cds\_10\_0\_chr22\_16287254\_r ) but different SNP IDs, meaning these lines represent 5 different SNPs that all overlap the same exon. Therefore we can find the total number of SNPs in an exon simply by counting the number of lines that have the same exon ID in the fourth column.

### ? Question

For the first 3 exons in your file, what is the number of SNPs that fall into that exon?

## Count the number of SNPs per exon

We've just seen how to count the number of SNPs in each exon, so let's do this for all the exons in our file.



## Hands-on: Counting SNPs

1. **Group** : Open the tool Group - data by a column and perform aggregate operation on other columns

### Settings

- o **Select data:** select dataset 3 (the output from the join tool)
- o **Group by column:** 4 (the column with the exon IDs)
- o **Insert operation:** click on this button, then set **Type** to `Count` and set **On column** to `Column: 4`

2. Make sure your screen looks like the image above and click **Execute** to perform the grouping. Your output dataset will look something like this:

4: Group on data 3	
3,897 lines	  
format: <b>tabular</b> , database: <a href="#">?</a>	
--Group by c4: count[c1]	
   	 
1	2
uc002zlh.1_cds_6_0_chr22_16277748_r	1
uc002zlj.1_cds_4_0_chr22_16277748_r	1
uc002zlj.1_cds_8_0_chr22_16287254_r	3
uc002zlp.1_cds_0_0_chr22_17071767_r	3
uc002zlv.3_cds_0_0_chr22_17264509_r	8
uc002zlv.3_cds_1_0_chr22_17280661_r	1

This file contains only two columns. The first contains the exon IDs, and the second the number of times that exon ID appeared in the file - in other words, how many SNPs were present in that exon.

### Question

How many exons are there in total in your file?

*Hint: Each line now represents a different exon, so you can see the answer to this when you expand the history item, as in the image above.*

## Sort the exons by SNPs count

Now we have a list of all exons and the number of SNPs they contain, but we would like to know which exons has the *highest number* of SNPs. We can do this by sorting the file on the second column.



## Hands-on: Sorting

1. **Sort** : Navigate to the tool `sort` - data in ascending or descending order
2. Set the **on column** parameter to `column: 2`, by default it will select a numerical sort in descending order, which is exactly what we want in this case.

The screenshot shows the Galaxy web interface. On the left, the 'Tools' panel is open, showing a list of available tools including 'sort'. The main workspace shows the 'Sort' tool configuration. The 'on column' parameter is highlighted with a red box and set to 'Column: 2'. The 'descending order' dropdown is also highlighted. The output history on the right shows a tabular database with 3,897 lines, grouped by column 4. The first few lines of the output are:

```
uc002zlh.1_cds_6_0_chr22_16277748_r 1
uc002zlj.1_cds_4_0_chr22_16277748_r 1
uc002xij.1_cds_8_0_chr22_16287254_r 3
uc002zlp.1_cds_0_0_chr22_17071767_r 3
uc002zlv.3_cds_0_0_chr22_17264509_r 8
uc002zlv.3_cds_1_0_chr22_17280661_r 1
```

3. Click **Execute** and examine the output file.

The screenshot shows the Galaxy output history for the 'Sort' tool. The title is '5: Sort on data 4'. It shows 3,897 lines in tabular format, grouped by column 4. The first few lines of the output are:

```
uc003bhh.3_cds_0_0_chr22_46652458_r 30
uc002zsw.2_cds_0_0_chr22_21044319_f 29
uc003alp.4_cds_5_0_chr22_32108069_r 17
uc010gqp.2_cds_10_0_chr22_16287254_r 17
uc003alo.2_cds_5_0_chr22_32108069_r 16
uc010gwj.1_cds_4_0_chr22_32108069_r 16
```

You should now see the same file as we had before, but the exons with the highest number of SNPs are now on top.

### Question

Which exon has the highest number of SNPs in your file?

Keep in mind this may depend on your settings when getting the data from UCSC.

## Select the top five exons

Let's say we want a list with just the top-5 exons with highest number of SNPs.



## Hands-on: Select first

1. **Select first** : Open the tool `select first - lines from a dataset`
2. Set **select first** to 5 and choose the sorted dataset from the previous step as the input.

select first

Get Data

Upload File from your computer

**Text Manipulation**

Select first lines from a dataset

Select last lines from a dataset

**Filter and Sort**

Filter data on any column using

Select first lines from a dataset (Galaxy Tool Version 1.0.0)

Select first

5

lines

from

5: Sort on data 4

Execute

1	2	3
uc003bh.3_cds_0_0_chr22_46652458_r	30	
uc002zsw.2_cds_0_0_chr22_21044319_f	29	
uc003alp.4_cds_5_0_chr22_32108069_r	17	
uc010gqp.2_cds_10_0_chr22_16287254_r	17	
uc003alp.2_cds_5_0_chr22_32108069_r	16	

3. Click **Execute** and examine the output file, this should contain only the first 5 lines of the previous dataset.

## Recovering exon info and displaying data in genome browsers

Congratulations! you have now determined which exons on chromosome 22 have the highest number of SNPs, but what else can we learn about them? One way to learn more about a genetic location is to view it in a genome browser. However, in the process of getting our answer, we have lost information about the location of these exons on the chromosome. But fear not, Galaxy saves all of your data, so we can recover this information quite easily.



## Hands-on: Compare two Datasets

1. **Compare two Datasets** : Open the tool compare two datasets - to find common or distinct rows
2. Set the parameters to compare the column 4 of the exon file with column 1 of the top-5 exons file to find matching rows.

3. Click **Execute** and examine your output file. It should contain the locations of your top 5 exons:

1	2	3	4	5	6
chr22	21044318	21045692	uc002zsw.2_cds_0_0_chr22_21044319_f	0	+
chr22	32108068	32113221	uc003alo.2_cds_5_0_chr22_32108069_r	0	-
chr22	32108068	32113277	uc003alp.4_cds_5_0_chr22_32108069_r	0	-
chr22	46652457	46659219	uc003bhh.3_cds_0_0_chr22_46652458_r	0	-
chr22	16287253	16287885	uc010gqp.2_cds_10_0_chr22_16287254_r	0	-

## UCSC genome browser

A good way to learn about these exons is to look at their genomic surrounding. This can be done by using genome browsers. Galaxy can launch a genome browser such as IGV on your local machine, and it can connect to online genome browsers as well. An example of such an online genome browser is the UCSC genome browser.

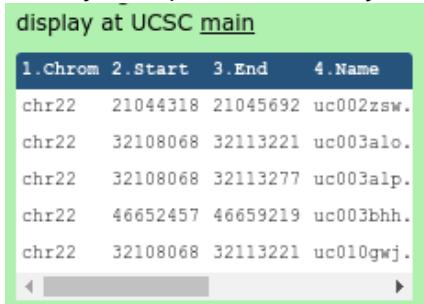


## Hands-on: UCSC genome browser

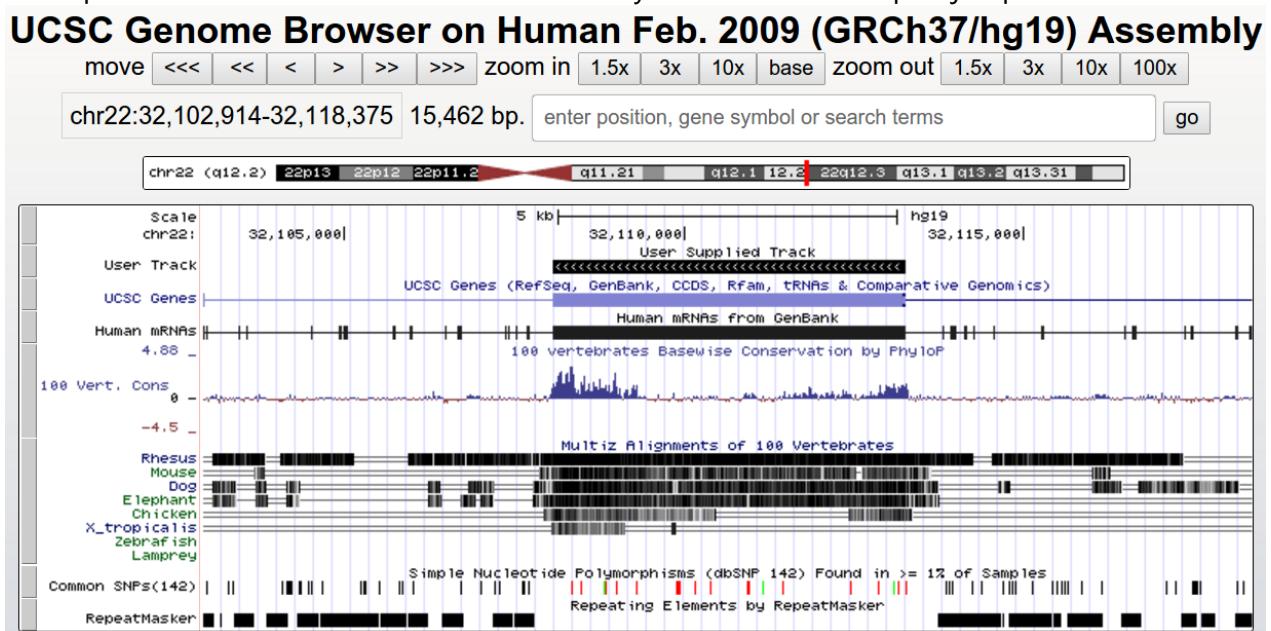
1. First, we have to tell Galaxy which **Genome build** this data uses (hg19), we can do this as follows:



2. To **visualize the data in UCSC genome browser**, click on **display at ucsc main** option visible when you expand the history item.



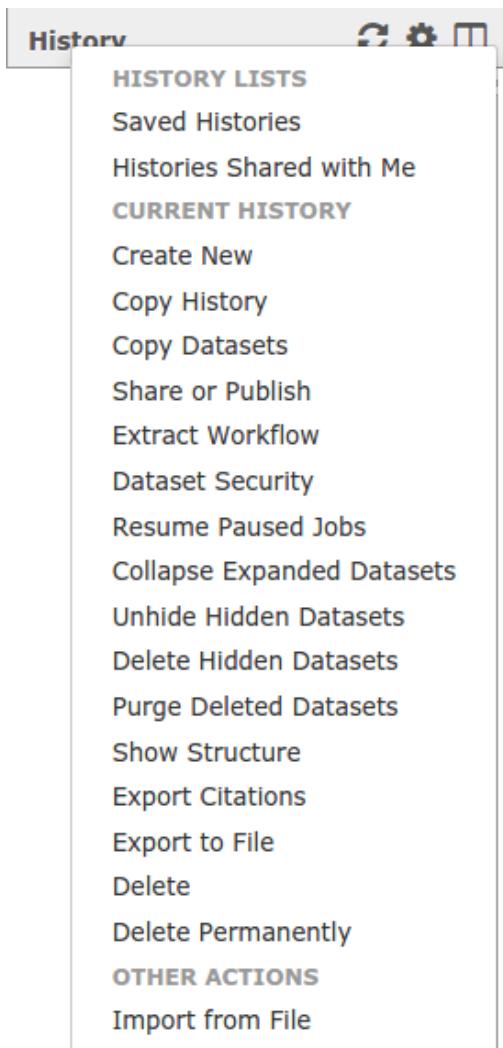
This will upload the data to UCSC as custom track. To see your data look at the **User Track** near the top. You can enter the coordinates of one of your exons at the top to jump to that location.



UCSC provides a large number of tracks that can help you get a sense of your genomic area, it contains common SNPs, repeats, genes, and much more (scroll down to find all possible tracks).

## Galaxy management

In Galaxy your analyses live in histories such as your current one. Histories can be very large, and you can have as many histories as you want. You can control your histories (switching, copying, sharing, creating a fresh history, etc.) in the **Options** menu on the top of the history pane (gear symbol):



If you create a new history, your current history does not disappear. If you would like to list all of your histories just choose `saved Histories` from the history menu and you will see a list of all your histories in the center pane:

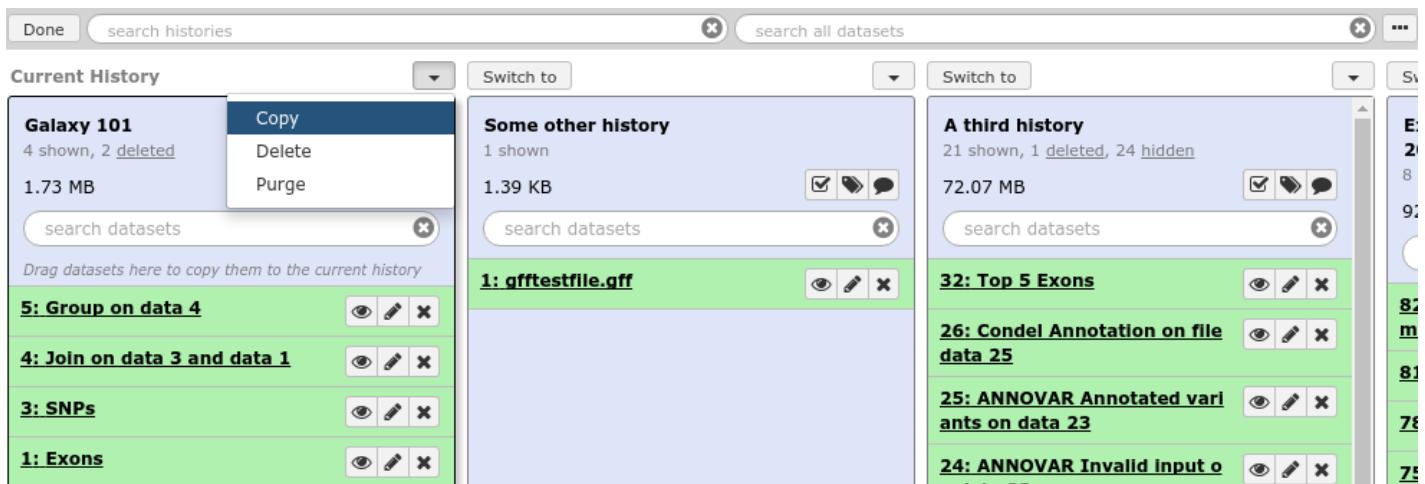
## Saved Histories

search history names and tags

[Advanced Search](#)

<input type="checkbox"/> <u>Name</u>	<b>Datasets</b>
<input type="checkbox"/> Galaxy 101 ▾	7

An alternative overview of your histories can be accessed by clicking on the **View all histories** button at top of your history pane (window icon).



The screenshot shows the Galaxy web interface with three main history panels. The left panel, titled 'Current History', contains a history named 'Galaxy 101' with 4 shown, 2 deleted datasets, totaling 1.73 MB. The middle panel, titled 'Some other history', contains a history with 1 shown dataset, totaling 1.39 KB. The right panel, titled 'A third history', contains a history with 21 shown, 1 deleted, 24 hidden datasets, totaling 72.07 MB. Each history panel has a 'Switch to' dropdown and a search bar. Individual datasets within the histories also have their own search bars and edit/delete icons.

Here you see a more detailed view of each history, and can perform the same operations, such as switching to a different history, deleting a history, purging it (permanently deleting it, this action cannot be reversed), or copying datasets and even entire histories.

You can always return to your analysis view by clicking on **Analyze Data** in the top menu bar.

## Convert your analysis history into a workflow

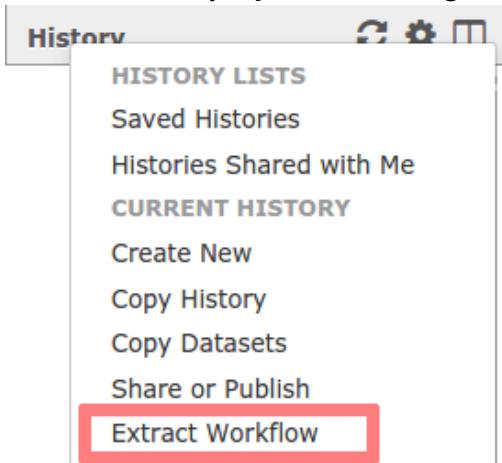
When you look carefully at your history, you can see that it contains all steps of our analysis, from the beginning to the end. By building this history we have actually built a complete record of our analysis with Galaxy preserving all parameter settings applied at every step. Wouldn't it be nice to just convert this history into a workflow that we'll be able to execute again and again?

Galaxy makes this very easy with the `Extract workflow` option. This means any time you want to build a workflow, you can just perform it manually once, and then convert it to a workflow, so that next time it will be a lot less work to do the same analysis.



## Hands-on: Extract workflow

1. **Clean up** your history. If you had any failed jobs (red), please remove those datasets from your history by clicking on the **x** button. This will make the creation of a workflow easier.
2. Go to the history **Options menu** (gear symbol) and select the `Extract workflow` option.



The center pane will change as shown below and you will be able to choose which steps to include/exclude and how to name the newly created workflow.

**Workflow name**

Find exons with highest number of SNPs

**Create Workflow****Check all****Uncheck all****Tool****History items created**

Unknown

*This tool cannot be used in workflows***1: Exons** Treat as input dataset

Unknown

*This tool cannot be used in workflows***2: SNPs** Treat as input dataset

Join

 Include "Join" in workflow**3: Join on data 2 and data 1**

Group

 Include "Group" in workflow**4: Group on data 3**

Sort

 Include "Sort" in workflow**5: Sort on data 4**

Select first

 Include "Select first" in workflow**6: Select first on data 5**

Compare two Datasets

 Include "Compare two Datasets" in workflow**7: Compare two Datasets on data 6 and data 1**

3. **Uncheck** any steps that shouldn't be included in the workflow (if any), and **rename** the workflow to something descriptive, for example `Find exons with the highest number of SNPs`.

4. Click on the **Create Workflow** button near the top.

You will get a message that the workflow was created. But where did it go?

5. Click on **Workflow** in the top menu of Galaxy. Here you have a list of all your workflows. Your newly created workflow should be listed at the top:

**Your workflows****Name**

Find exons with highest number of SNPs ▾

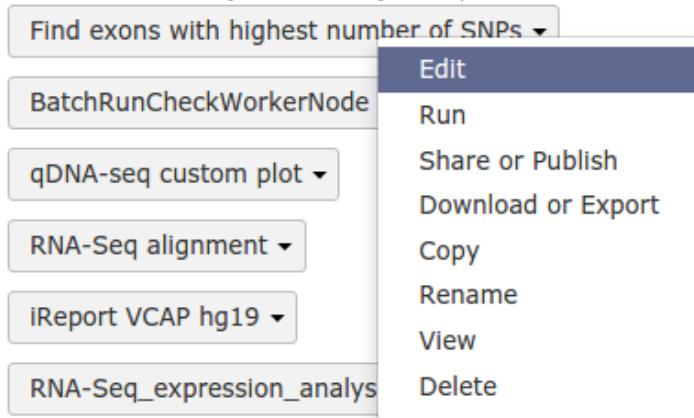
# The workflow editor

We can examine the workflow in Galaxy's workflow editor. Here you can view/change the parameter settings of each step, add and remove tools, and connect an output from one tool to the input of another, all in an easy and graphical manner. You can also use this editor to build workflows from scratch.

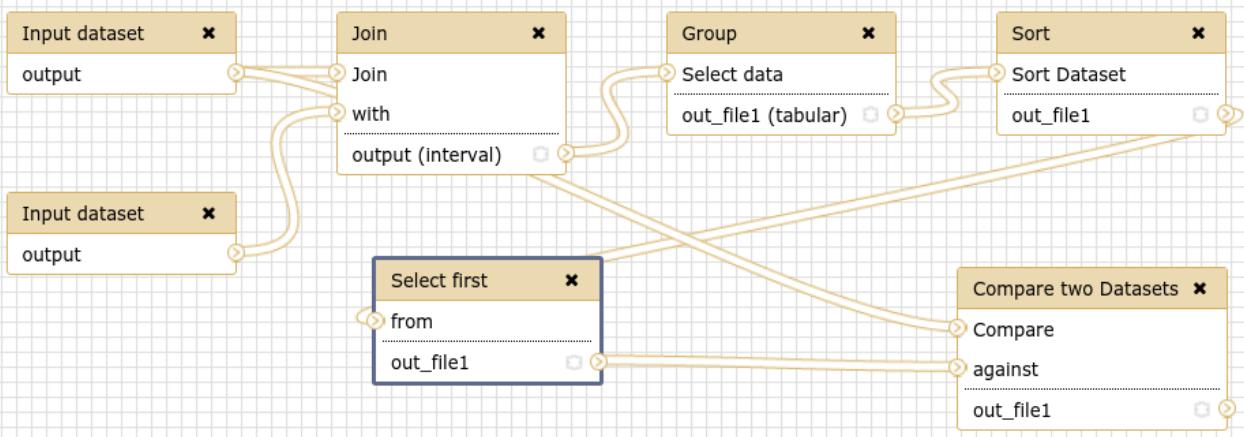


## Hands-on: Extract workflow

1. Click on the triangle to the right of your workflow name.



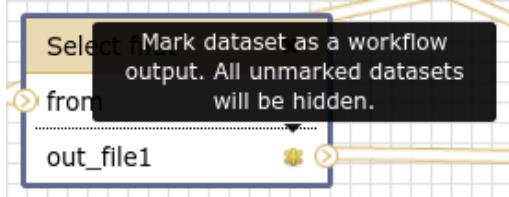
2. Select **Edit** to launch the workflow editor. You should see something like this:



When you click on a component, you will get a view of all the parameter settings for that tool on the right-hand side of your screen.

### Tip: Hiding intermediate steps

When a workflow is executed, the user is usually primarily interested in the final product and not in all intermediate steps. By default all the outputs of a workflow will be shown, but we can explicitly tell Galaxy which output to show and which to hide for a given workflow. This behaviour is controlled by the little asterisk next to every output dataset:



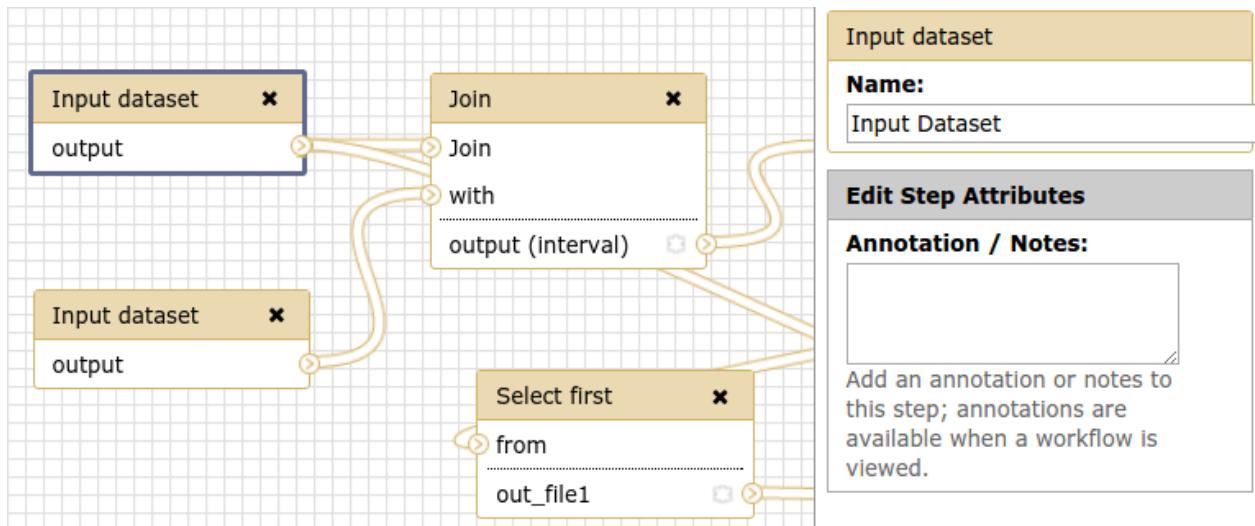
If you click on this asterisk for any of the output datasets, then *only* files with an asterisk will be shown, and all outputs without an asterisk will be hidden. (Note that clicking *all* outputs has the same effect as clicking *none* of the outputs, in both cases all the datasets will be shown.)

3. Click the asterisk next to `out_file1` in the `Select First` and `Compare two Datasets` tools.

Now, when we run the workflow, we will only see the final two outputs, our list with the top-5 exons and their SNP counts, and the file with exons ready for viewing in a genome browser. Once you have done this, you will notice that the **minimap** at the bottom-right corner of your screen will have a colour-coded view of your workflow, with orange boxes representing a tool with an output that will be shown.



If you didn't specify a name for the input files at the beginning they will be labeled `Input dataset`. In this case you can rename them now to avoid confusion when using the workflow later on.



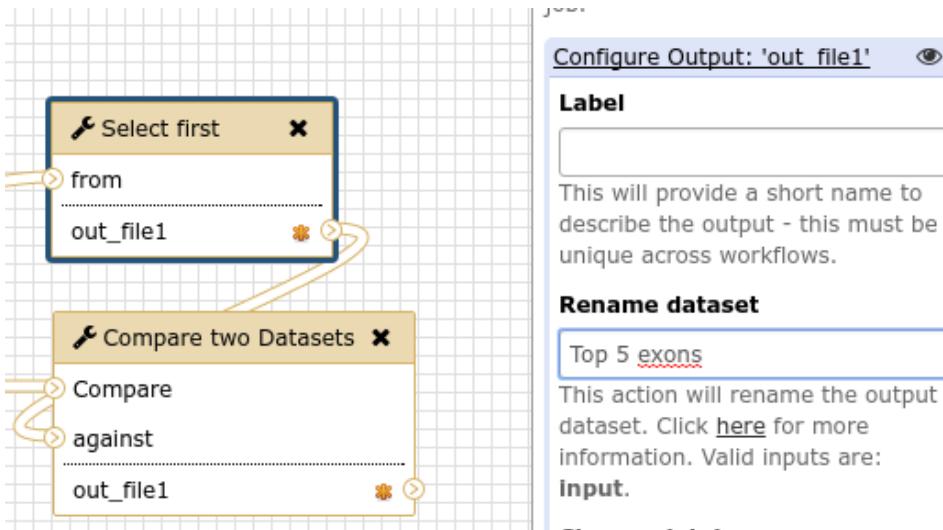
In the image above, you see that the top input dataset (with the blue border), connects to the first input of the join tool, so this corresponds to the exon data.

4. Click on the box corresponding to the exon input dataset, and **rename** it to `Exons` on the right-hand side of your screen.

Input dataset
<b>Name:</b>
Exons

5. **Repeat** this process for the other input dataset. Name it `Features`. We used it to calculate highest number of SNPs, but this workflow would also work with other features, so we give it a bit more generic name.

6. Let's also **rename the outputs**. Click on the `Select first` tool and in the menu on the right click on `configure output` and enter a descriptive name for the output dataset in the `Rename dataset` box.



7. **Repeat** this for the output of the `Compare two Datasets` tool.
8. **Save your workflow** (important!) by clicking on the gear icon at the top right of the screen, and selecting `Save`.



9. **Return** to the analysis view by clicking on `Analyze Data` at the top menu bar.

### Comments

We could **validate** our newly built workflow by running it on the same input datasets than the ones in the `Galaxy 101` history used to extract the workflow in order to make sure we do obtain the same results.

## Run workflow on different data

Now that we have built our workflow, let's use it on some different data. For example, let's find out which exons have the highest number of repeat elements.



## Hands-on: Run workflow

1. Create a **new history** (gear icon) and give it a name.
2. We will need the list of exons again. We don't have to get this from UCSC again, we can just **copy** it from our previous history. The easiest way to do this is to go to the history overview (window icon at top of history pane). Here you can just drag and drop datasets from one history to another.

The screenshot shows the Galaxy web interface with two history panes. The left pane is titled 'Current History' and contains a single dataset named '1: Exons'. A tooltip 'This history is empty' is displayed. The right pane is titled 'Galaxy 101' and contains a list of workflow steps:

- 21: Compare two Datasets on d and data 1
- 20: Select first on data 19
- 19: Sort on data 18
- 18: Group on data 17
- 17: Join on data 2 and data 1
- 2: SNPs
- 1: Exons

Each step in the Galaxy 101 history has a set of icons for eye, edit, and delete.

3. We wanted to know something about the repetitive elements per exon. We get this data from UCSC.
  - o **assembly** should be set to Feb. 2009 (GRCh37/hg19)
  - o **group** parameter should be `Repeats`
  - o **position** should be `chr22`
  - o leave the rest of the settings to the defaults

Click on `Get output` and then `send query to Galaxy` on the next screen.

4. Open the **workflow menu** (top menu bar). Find the workflow you made in the previous section, and select the option `Run`.

## Your workflows

### Name

Find exons with highest number of SNPs

Edit

Run

Share or Publish

Download or Export

Copy

Rename

View

Delete

The center pane will change to allow you to configure and launch the workflow.

5. Select appropriate datasets for the inputs as shown below, then scroll down and click **Run workflow**.

Step 1: Input dataset

**exons** 

1: Exons 

type to filter

Step 2: Input dataset

**features** 

2: UCSC Main on Human: rmsk (chr22:1-51304566) 

type to filter

Step 3: Join (version 1.0.0)

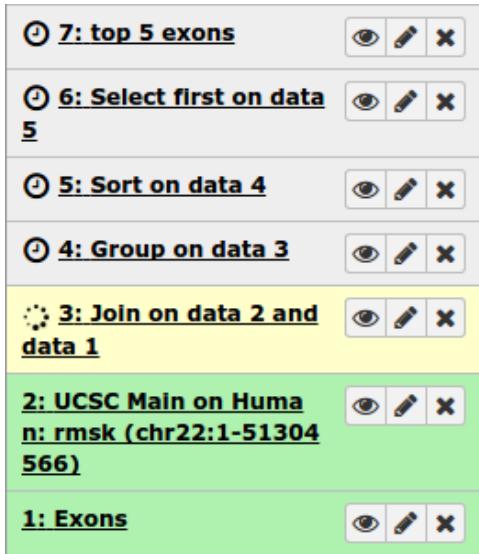
Step 4: Group (version 2.1.0)

Step 5: Sort (version 1.0.3)

Step 6: Select first (version 1.0.0)

Step 7: Compare two Datasets (version 1.0.2)

Once the workflow has started you will initially be able to see all its steps:



### Comment

Because most intermediate steps of the workflow were hidden, once it is finished you will only see the final two datasets. If we want to view the intermediate files after all, we can unhide all hidden datasets by selecting `Include Hidden datasets` from the history options menu.

### Questions

Which exon had the highest number of repeats? How many repeats were there?

## Share your work

One of the most important features of Galaxy comes at the end of an analysis. When you have published striking findings, it is important that other researchers are able to reproduce your in-silico experiment. Galaxy enables users to easily share their workflows and histories with others.

To share a history, click on the gear symbol in the history pane and select `share` or `Publish`. On this page you can do 3 things:

1. **Make accessible via Link.** This generates a link that you can give out to others. Anybody with this link will be able to view your history.
2. **Publish History.** This will not only create a link, but will also publish your history. This means your history will be listed under `shared Data` → `Published Histories` in the top menu.
3. **Share with Individual Users.** This will share the history only with specific users on the Galaxy instance.



### Hands-on: Share history and workflow

1. Share one of your histories with your neighbour.
2. See if you can do the same with your workflow!
3. Find the history and/or workflow shared by your neighbour. Histories shared with specific users can be accessed by those users in their history menu (gear icon) under Histories shared with me .

## Conclusion

🎉 Well done! 🙌 You have just performed your first analysis in Galaxy. You also created a workflow from your analysis so you can easily repeat the exact same analysis on other datasets. Additionally you shared your results and methods with others.



### Key points

- Galaxy provides an easy-to-use graphical user interface for often complex commandline tools
- Galaxy keeps a full record of your analysis in a history
- Workflows enable you to repeat your analysis on different data
- Galaxy can connect to external sources for data import and visualization purposes
- Galaxy provides ways to share your results and methods with others

👏 Congratulations on successfully completing this tutorial!



### Feedback

Please take a moment and provide your feedback on this tutorial. Your feedback will help guide and improve future revisions to this tutorial. Feedback Form (<https://tinyurl.com/GTNfeedback>)

*This material is the result of a collaborative work. Thanks to all the contributors (<https://galaxyproject.github.io/training-material/introduction#contributors>) and the Galaxy Training Network (<https://wiki.galaxyproject.org/Teach/GTN>)!*

*Found a typo? Something is wrong in this tutorial? Edit it on GitHub (<https://github.com/galaxyproject/training-material/tree/master/introduction/tutorials/galaxy-intro-101.md>).*